

# Utilization of Antibiograms

**Joe Sartor, Pharm D**

A series of horizontal lines of varying lengths and colors (teal, light blue, white) extending from the right side of the slide towards the center.

# The History of Medicine

HAVE EARACHE?  
EAT ROOT.  
2000 BC



That root is heathen.  
Say this prayer.  
1000 AD



That prayer is  
superstition.  
Drink this potion.  
1700 AD



That potion is snake oil.  
Swallow these pills.  
1900 AD



Those pills are placebo.  
Take antibiotics and  
get immunized.  
1960 AD



Antibiotics are artificial.  
Immunizations cause autism.  
Eat this root.  
2010 AD

J. Chang MD

PoorMD.com

# Is There A Need for Antibiogram?

## Antimicrobial resistance

New antibiotics to cover Gram+ organisms ,  
MRSA, VRE

telavancin (Vibativ)

oritavancin (Orbactiv)

dalbavancin (Dalvance)

ceftaroline (Teflaro)

daptomycin (Cubicin)

tigecycline (Tygacil)

tedizolid (Sivextro)

# Is There A Need for Antibigram?

**New antibiotics to cover gram- organisms'**  
**ceftazidime/avibactam (Avycaz)**  
**Ceftolozane/tazobactam (Zerbaxa)**

# Is There A Need for Antibigram?

- **Increasing resistance is often associated with inappropriate therapy, esp. empiric therapy**
- **Inappropriate therapy – Increased mortality, increased LOS**
- **Clinical outcomes – increased morbidity, increased mortality**

# The Antibiogram

- **Antibiogram can be utilized to aid in appropriate selection of empiric therapy**
- **Provides susceptibility rates to optimize empiric therapy – increases probability of initiating appropriate empiric therapy**
- **Aids the making of clinical decisions, infection control interventions, resistance control**

# The Antibiogram

- Susceptibility of pathogens to commonly used antimicrobials
- Data from individual susceptibility reports of individual pathogens
- CLSI Guidelines are critical to standardized isolate selections and susceptibility testing and reporting
- Generated by clinical microbiology laboratory
- Can be used by any health care professional involved in prevention and treatment of infectious disease

# Data analysis for generating antibiogram - CLSI Guidelines

- N = 30
- Once yearly required
- Multiple institutions , may include clinics, other institutions using lab services - **not recommended**
- Can include all specimen types, but may segregate (urine/non-urine )
- May include variety of patient types and settings, data stratification, infection site and type
- Report % susceptible only
- Report in a format easily accessible to clinicians



# Data analysis for generating antibiogram

Frequency once a year more frequent if

- Large number of isolates
- New antimicrobial agents
- Clinically important changes have occurred or are perceived
- Seasonal variations in resistance
- Small sample of isolates

Isolates include first isolate of a given species/patient analysis period, organism with > 30 isolates, isolates collected for diagnosis purposes should be included

- Do not include duplicate isolates from the same patient or isolates from surveillance cultures, environmental cultures or other non-patient sources

# Generalities from Antibioqram

High Nosocomial MRSA=poor infection control

High VRE rates may indicate over-utilization of  
Vancomycin particularly oral dosing

ESBL rates might indicate over-utilization of  
cephalosporins/penicillins

High KPC rates = over use of  
cephalosporins/carbapenems

# Pitfalls of antibiogram

- **Small Sample = 30 isolates minimum**
- **Multiple institutions = ??????????????**
- **Updated at least annually, with large number of isolates more frequently, if more frequent are things changing**
- **May include variety of patient types and settings, data stratification, infection site and type**
- **Is break point for susceptible organisms optimal  
Vancomycin, Piperacillin/tazobactam**
- **Selection of combination therapy to cover resistance**

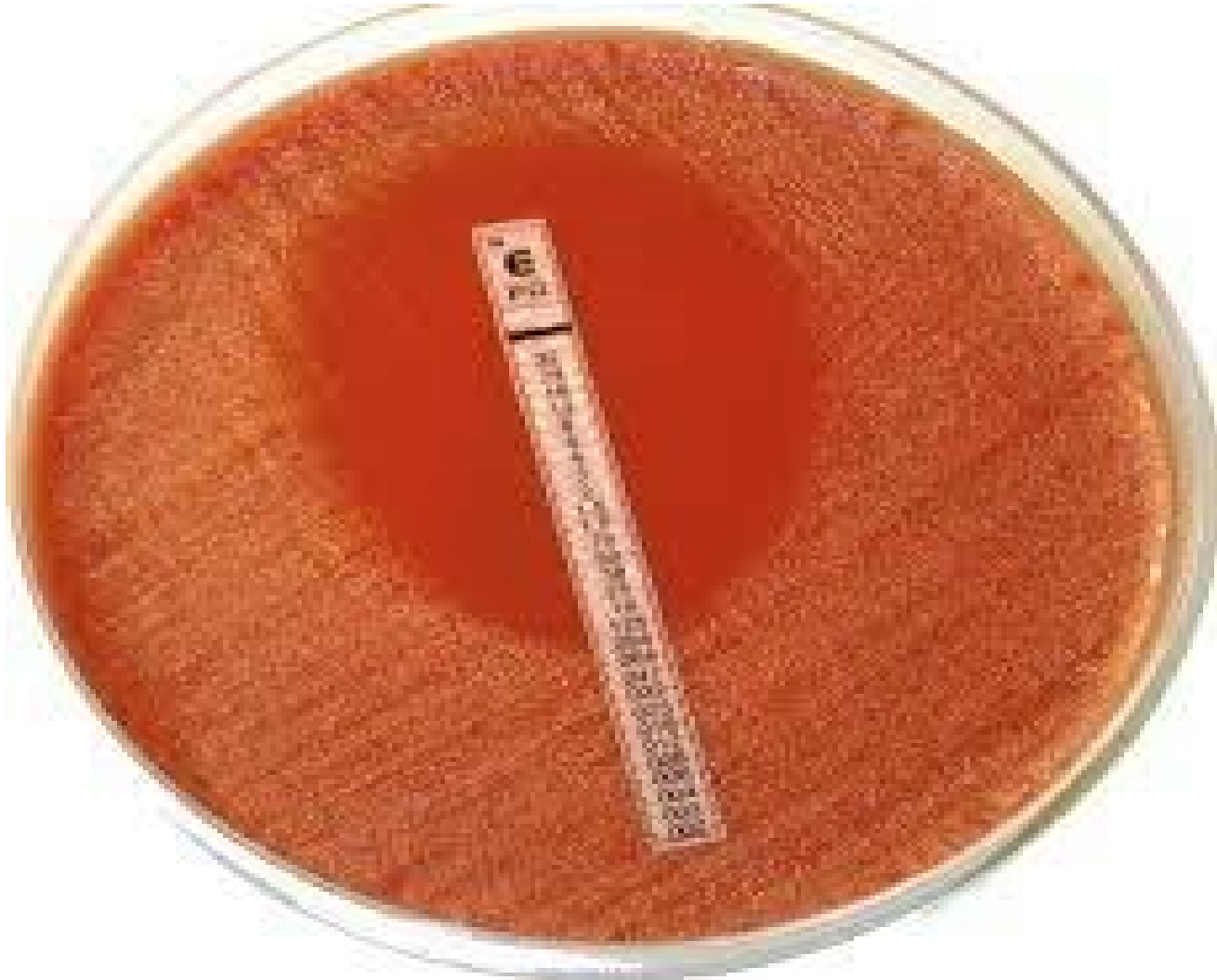
# Reading the antibiogram

- n= standards recommend including only the first isolate/patient for analysis = number of patients with pathogen.
- Repeat admissions of same patient might dilute results
- Which pathogens are most common >n
- Pseudomonas doesn't usually have one antibiotic with excellent activity
- ESBL - cefepime is good marker for ESBL gram-pathogens
- KPC – resistant to carbapenems and all other b-lactams approximation looking at imipenem.
- MRSA %
- VANCOMYCIN usually 100% but MIC is important 2mcg/ml
- Enterococcus faecium vs faecalis

# Etest



# Etest, Epsilon meter test



# Etest

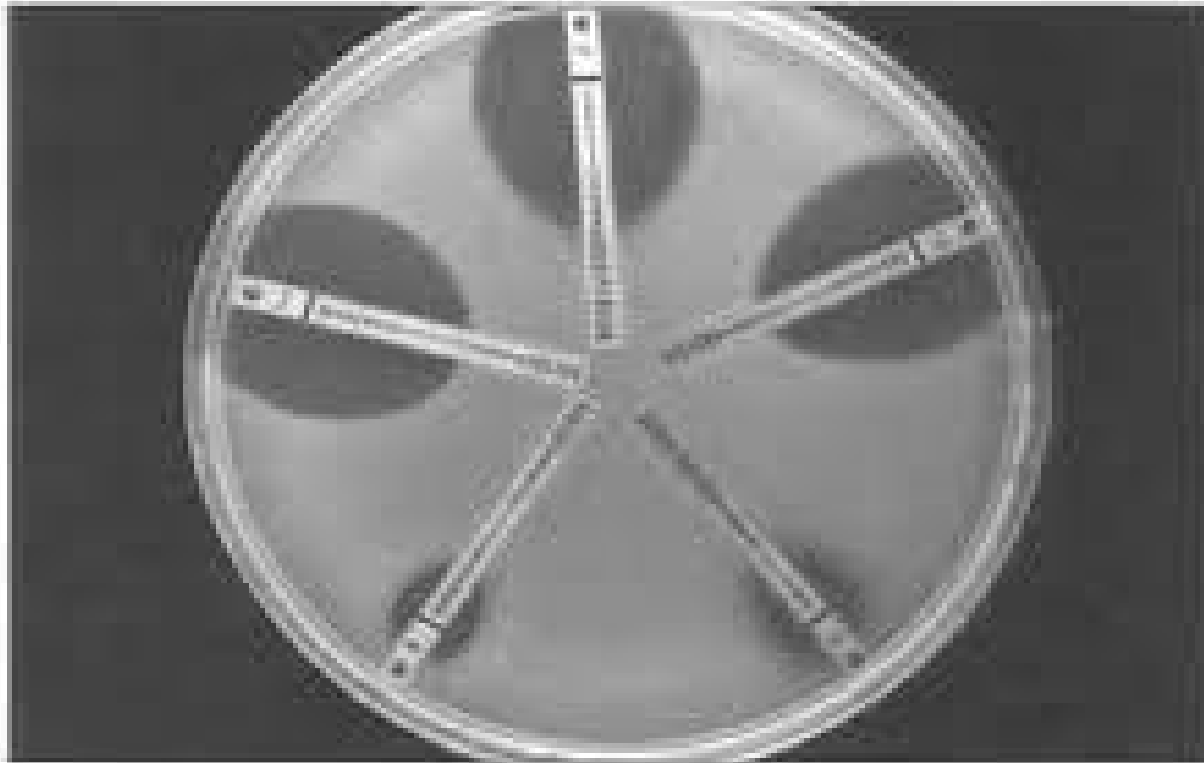


Figure 1-Photograph of a 15-centimeter long Mueller-Hinton plate with five E tests strips (ciprofloxacin, ceftazidime, piperacillin, ticarcillin-clavulanic acid and trimethoprim/ sulfamethoxazole). The microorganism being tested was *Xanthoma maltophilia*.

**UT HEALTH NORTHEAST  
DEPARTMENT OF PATHOLOGY - MICROBIOLOGY  
2015 NON-URINE ANTIBIOGRAM**

ORGANISM	# ISOLATES	% SUSCEPTIBLE																			
		ANTIMICROBIC																			
		A/S	AK	AM	AZT	CAX	CAZ	CFT	CFZ	CP	CPE	CRM	ETP	GM	IMP	LVX	MER	P/T	T/S	TGC	TO
<b>GRAM NEGATIVE</b>																					
Acinetobacter sp.	88	83	91			65	84	52		80	80			85		85	94		86		86
Achromobacter sp.	37		64		17	44	50	28		44	47			67	92	72	89	78	86		61
Enterobacter sp.	163	48	98	22	88	83	93	90	21	98	97	47	99	93	94	99	99	95	93	98	94
E. coli	135	45	99	40	93	93	93	94	83	64	94	91	100	84	99	64	100	97	59	100	84
Klebsiella sp.	210	81	99<1		97	98	98	98	84	99	99	92	99	97	100	99	100	99	93	99	97
Proteus sp.	58	74	98	62	90	93	95	95	69	71	95	90	100	95	50	84	100	100	67	100	95
Pseudomonas sp.	600		87		66	50	90	29		87	85			76	86	87	92	95	62		92
Serratia sp.	162	15	99	8	69	60	65	70<1		94	97	1	100	94	95	97	100	57	97	99	86
Stenotrophomonas sp.	244															91				99	

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<b>GRAM POSITIVE</b>																															
Enterococcus sp.	60		95				0					70			97	12		77		75	98					95	45	70		17	95
MRSA	261	0		0		82	100	68			0	35			99	12	98		0	38	99	0	70	0	0	95			99	99	100
MSSA	374	99	0	100		89	54	80			100	86			99	66	99		100	89	98	100	95	100	19	96		99	99	95	100
Coag Neg Staph	155	54	0	54		98	68					65			97	38	88			66	99		78	54	17	99		98	64	82	99
Strep pneumoniae	42			98	63	95		83	71	98			95	80		63					100	93				71			76	78	100

Data collected 01/01/15 through 12/31/15

41.0% of the Staph aureus isolated were MRSA

MRSA = Methicillin Resistant Staph aureus

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**% SUSCEPTIBLE**

ORGANISM	# ISOLATES	ANTIMICROBIC	Amp/ Sulbactam	Amikacin	Ampicillin	Aztreonam	Ceftriaxone	Ceftazidime	Cefotaxime	Cefazolin	Ciprofloxacin	Cefepime	Cefuroxime	Ertapenem	Nitrofurantoin	Gentamicin	Imipenem	Levofloxacin	Merepenem	Piperacilin/Tazo	Trimeth/Sulfa	Tigecycline	Tobramycin
GRAM NEGATIVE			A/S	AK	AM	AZT	CAX	CAZ	CFT	CFZ	CP	CPE	CRM	ETP	FD	GM	IMP	LVX	MER	P/T	T/S	TGC	TO
E. coli	589		58	99	53	96	96	96	97	88	78	97	91	100	98	93	100	78	100	96	68	100	94
Klebsiella sp.	156		84	100	0	96	96	96	96	88	96	96	92	99	49	98	100	97	100	96	92	99	97
Proteus sp.	69		78	99	74	90	90	94	93	81	84	94	91	100	0	88	0	86	100	100	80	100	90
Pseudomonas sp.	43			95		77	50	98	33		84	93				81	84	81	93	100	33		95

ORGANISM	# ISOLATES	ANTIMICROBIC	Amp/sulb	Ampicillin	Augmentin	Ceftriaxone	Ciprofloxacin	Daptomycin	Nitrofurantoin	Gentamicin	Gentamicin Synergy	Levofloxacin	Linezolid	Oxacillin	Penicillin	Rifampin	Strep Synergy	Synercid	Trimeth/Sulf	Tetracycline	Vancomycin
GRAM POSITIVE			A/S	AM	AUG	CAX	CP	DAP	FD	GM	GMS	LVX	LZD	OX	P	RIF	STS	SYN	T/S	TE	VA
Coag Neg Staph	76		33	0	33	33	54	100	97	88		54	100	33	14	99		100	71	86	100
Enterococcus sp.	141			90			58	98	94		70	63	94		89	37	70			17	91
Strep agalactiae	13							100				100	100		100						100
MRSA	0																				
MSSA	23		23	0	100	100	83	96	100	100		87	100	100	17	100		100	100	96	100

Data collected 01/01/15 thru 12/31/15

No MRSA isolated from urine in 2015

# Fluroquinolone use is associated with:

- Increased risk of *Clostridium difficile* NAP1/027 hypervirulent/epidemic strain
- Increased risk of vancomycin-resistant *Enterococcus*
- Increased risk of ESBL *Enterobacteriaceae* (*E coli*, *Klebsiella*)
- Increased *Pseudomonas* meropenem resistance
- Increased Carbapenem Resistant *Enterobacteriaceae*

1. Center for Infection Disease ANTIBIOTIC RESISTANCE THREATS in the United States, 2013

2. Hayakawa et al. January 2013 Volume 57 Number 1 Antimicrobial Agents and Chemotherapy p. 49–55

3. Rodriguez-Bano J, Navarro MD, Romero L, Muniain MA, Perea EJ, Perez-Cano R, et al Clin Infect Dis 2006;42(1):37-45.

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MSSA	374	99	0	100		89	54	80			100	86			99	66	99		100	89	98	100	95	100	19	96		99	99	95	100
Coag Neg Staph	155	54	0	54			98	68				65			97	38	88			66	99		78	54	17	99					99
Strep pneumoniae	42			98	63	95		83	71	98			95	80		63				100		93			71				76	78	100

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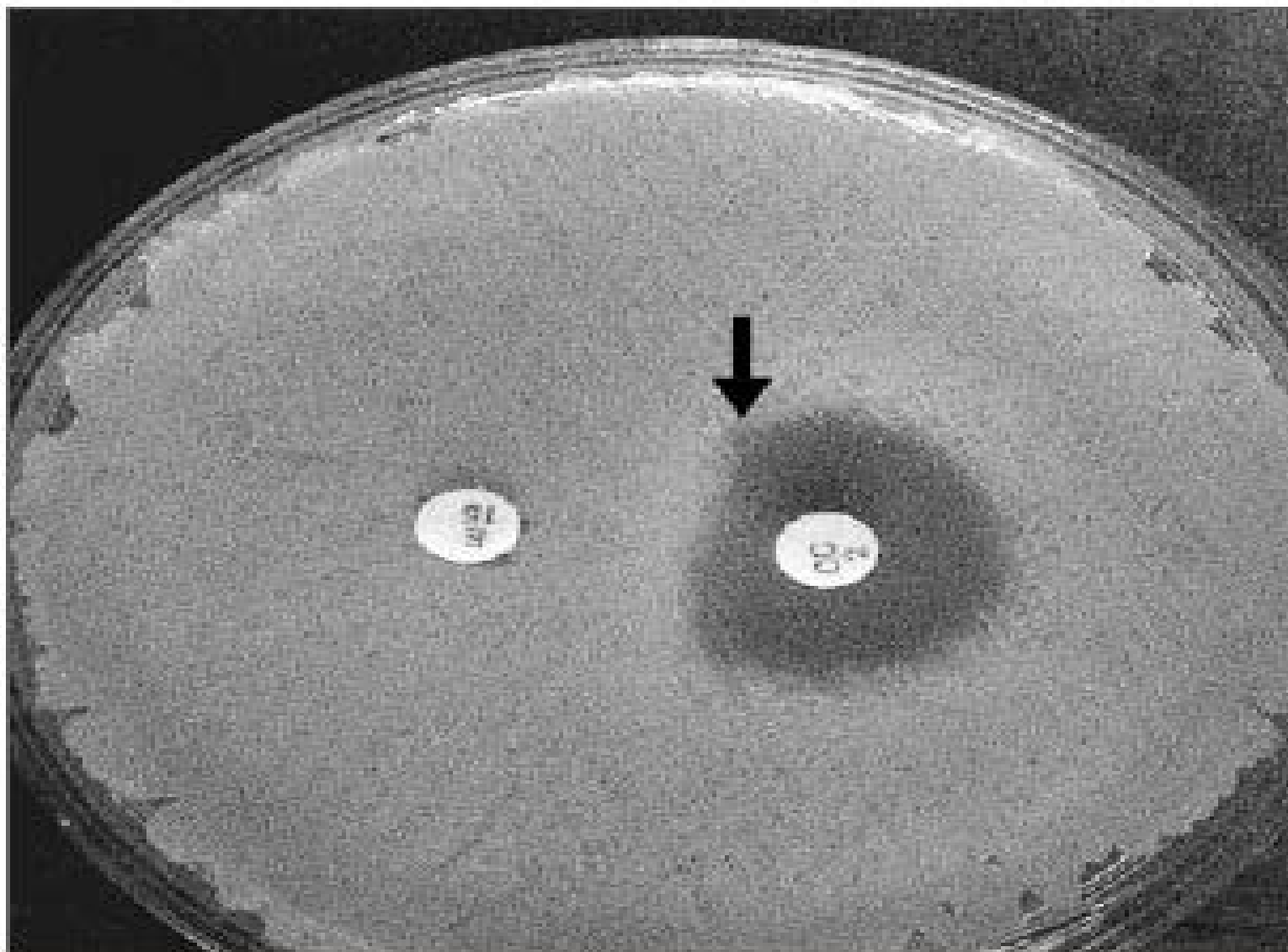
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MSSA = Methicillin Susceptible Staph aureus



**Figure 3.** The D-Zone Test for Erythromycin-Resistant, Clindamycin-Susceptible Isolates.



**UT HEALTH NORTHEAST  
DEPARTMENT OF PATHOLOGY - MICROBIOLOGY  
2015 NON-URINE ANTIBIOGRAM**

ORGANISM	# ISOLATES	% SUSCEPTIBLE																				
		ANTIMICROBIC	A/S	AK	AM	AZT	CAX	CAZ	CFT	CFZ	CP	CPE	CRM	ETP	GM	IMP	LVX	MER	P/T	T/S	TGC	TO
Acinetobacter sp.	88		83	91		65	84	52		80	80				85		85	94		86		86
Achromobacter sp.	37			64	17	44	50	28		44	47			67	92	72	89	78	86			61
Enterobacter sp.	163		48	98	22	88	83	93	90	21	98	97	47	99	93	94	99	99	95	93	98	94
E. coli	135		45	99	40	93	93	93	94	83	64	94	91	100	84	99	64	100	97	59	100	84
Klebsiella sp.	210		81	99<1		97	98	98	98	84	99	99	92	99	97	100	99	100	99	93	99	97
Proteus sp.	58		74	98	62	90	93	95	95	69	71	95	90	100	95	50	84	100	100	67	100	95
Pseudomonas sp.	600			87	66	50	90	29		87	85			76	86	87	92	95	62			92
Serratia sp.	162		15	99	8	69	60	65	70<1	94	97	1	100	94	95	97	100	57	97	99	86	
Stenotrophomonas sp.	244							38								91				99		

ORGANISM	# ISOLATES	% SUSCEPTIBLE																														
		ANTIMICROBIC	A/S	AM	AUG	AZI	C	CAX	CD	CFR	CFT	CFZ	CP	CPE	CRM	DAP	E	GM	GMS	IMP	LVX	LZD	MER	MXF	OX	P	RIF	STS	SYN	T/S	TE	VA
Enterococcus sp.	60		95				0					70			97	12		77		75	98				95	45	70				17	95
MRSA	261		0	0		82	100	68			0	35			99	12	98		0	38	99	0	70	0	0	95		99	99	95	100	
MSSA	374		99	0	100	89	54	80			100	86			99	66	99		100	89	98	100	95	100	19	96		99	99	95	100	
Coag Neg Staph	155		54	0	54			68				65			97	38	88			66	99		78	54	17	99		98	64	82	99	
Strep pneumoniae	42				98	63	95		83	71	98		95	80		63					100		93		71			76	78	100		

Data collected 01/01/15 through 12/31/15  
41.0% of the Staph aureus isolated were MRSA

MRSA = Methicillin Resistant Staph aureus

MSSA = Methicillin Susceptible Staph aureus

**UT HEALTH NORTHEAST  
DEPARTMENT OF PATHOLOGY - MICROBIOLOGY  
2015 NON-URINE ANTIBIOGRAM**

ORGANISM	# ISOLATES	% SUSCEPTIBLE																			
		ANTIMICROBIC																			
GRAM NEGATIVE		A/S	AK	AM	AZT	CAX	CAZ	CFT	CFZ	CP	CPE	CRM	ETP	GM	IMP	LVX	MER	P/T	T/S	TGC	TO
Acinetobacter sp.	88	83	91			65	84	52		80	80			85		85	94			86	86
Achromobacter sp.	37		64		17	44	50	28		44	47			67	92	72	89	78	86		61
Enterobacter sp.	163	48	98	22	88	83	93	90	21	98	97	47	99	93	94	99	99	95	93	98	94
E. coli	135	45	99	40	93	93	93	94	83	64	94	91	100	84	99	64	100	97	59	100	84
Klebsiella sp.	210	81	99	<1	97	98	98	98	84	99	99	92	99	97	100	99	100	99	93	99	97
Proteus sp.	58	74	98	62	90	93	95	95	69	71	95	90	100	95	50	84	100	100	67	100	95
Pseudomonas sp.	600		87		66	50	90	29		87	85			76	86	87	92	95	62		92
Serratia sp.	162	15	99	8	69	60	65	70	<1	94	97	1	100	94	95	97	100	57	97	99	86
Stenotrophomonas sp.	244						38									91				99	

ORGANISM	# ISOLATES	ANTIMICROBIC																													
		A/S	AM	AUG	AZI	C	CAX	CD	CFR	CFT	CFZ	CP	CPE	CRM	DAP	E	GM	GMS	IMP	LVX	LZD	MER	MXF	OX	P	RIF	STS	SYN	T/S	TE	VA
GRAM POSITIVE		A/S	AM	AUG	AZI	C	CAX	CD	CFR	CFT	CFZ	CP	CPE	CRM	DAP	E	GM	GMS	IMP	LVX	LZD	MER	MXF	OX	P	RIF	STS	SYN	T/S	TE	VA
Enterococcus sp.	60		95				0					70			97	12		77		75	98				95	45	70			17	95
MRSA	261	0		0		82	100	68			0	35			99	12	98		0	38	99	0	70	0	0	95		99	99	95	100
MSSA	374	99	0	100		89	54	80			100	86			99	66	99		100	89	98	100	95	100	19	96		99	99	95	100
Coag Neg Staph	155	54	0	54		98	68					65			97	38	88			66	99		78	54	17	99		98	64	82	99
Strep pneumoniae	42			98	63	95		83	71	98			95	80		63				100		93			71				76	78	100

Data collected 01/01/15 through 12/31/15  
41.0% of the Staph aureus isolated were MRSA

MRSA = Methicillin Resistant Staph aureus

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**UT HEALTH NORTHEAST  
DEPARTMENT OF PATHOLOGY - MICROBIOLOGY  
2015 NON-URINE ANTIBIOGRAM**

ORGANISM	# ISOLATES	% SUSCEPTIBLE																			
		ANTIMICROBIC																			
GRAM NEGATIVE		A/S	AK	AM	AZT	CAX	CAZ	CFT	CFZ	CP	CPE	CRM	ETP	GM	IMP	LVX	MER	P/T	T/S	TGC	TO
Acinetobacter sp.	88	83	91			65	84	52		80	80			85		85	94			86	86
Achromobacter sp.	37		64		17	44	50	28		44	47			67	92	72	89	78	86		61
Enterobacter sp.	163	48	98	22	88	83	93	90	21	98	97	47	99	93	94	99	99	95	93	98	94
E. coli	135	45	99	40	93	93	93	94	83	64	94	91	100	84	99	64	100	97	59	100	84
Klebsiella sp.	210	81	99	<1	97	98	98	98	84	99	99	92	99	97	100	99	100	99	93	99	97
Proteus sp.	58	74	98	62	90	93	95	95	69	71	95	90	100	95	50	84	100	100	67	100	95
Pseudomonas sp.	600		87		66	50	90	29		87	85			76	86	87	92	95	62		92
Serratia sp.	162	15	99	8	69	60	65	70	<1	94	97	1	100	94	95	97	100	57	97	99	86
Stenotrophomonas sp.	244						38									91				99	

ORGANISM	# ISOLATES	ANTIMICROBIC																													
		A/S	AM	AUG	AZI	C	CAX	CD	CFR	CFT	CFZ	CP	CPE	CRM	DAP	E	GM	GMS	IMP	LVX	LZD	MER	MXF	OX	P	RIF	STS	SYN	T/S	TE	VA
GRAM POSITIVE																															
Enterococcus sp.	60		95				0				70			97	12		77		75	98					95	45	70			17	95
MRSA	261	0		0		82	100	68			0	35		99	12	98		0	38	99	0	70	0	0	95		99	99	95	100	
MSSA	374	99	0	100		89	54	80			100	86		99	66	99		100	89	98	100	95	100	19	96		99	99	95	100	
Coag Neg Staph	155	54	0	54		98	68				65			97	38	88			66	99		78	54		17	99				99	
Strep pneumoniae	42			98	63	95		83	71	98		95	80		63					100		93			71				76	78	100

Data collected 01/01/15 through 12/31/15

41.0% of the Staph aureus isolated were MRSA

MRSA = Methicillin Resistant Staph aureus

MSSA = Methicillin Susceptible Staph aureus

**UNIVERSITY OF TEXAS HEALTH SCIENCE CENTER TYLER  
DEPARTMENT OF PATHOLOGY - MICROBIOLOGY  
2010 NON-URINE ANTIBIOGRAM  
% SUSCEPTIBLE**

ORGANISM	# ISOLATES	ANTIMICROBIC																											
		Amp/Sulbactam	Amikacin	Ampicillin	Augmentin	Aztreonam	Ceftioxone	Ceftazidime	Ceftaxime	Cefazolin	Ciprofloxacin	Cefepime	Cefturoxime	Cefotetan	Gentamicin	Imipenem	Levofloxacin	Meropenem	Moxifloxacin	Pip/Tazobactam	Trim/Sulfa	Ticarc/Clavulanate	Tobramycin						
<b>GRAM NEGATIVE</b>		A/S	AK	AM	AUG	AZT	CAX	CAZ	CFT	CFZ	CP	CPE	CRM	CTN	GM	IMP	LVX	MER	MXF	P/T	T/S	TIM	TO						
Acinetobacter sp.	36	91	94			84	79	84			88	82			88		97	91			94	91	88						
Achromobacter sp.	5		80			40	40	100	80		80	80			80	100	100	100		100	100	80	80						
Enterobacter sp.	52	54	100	25		88	92	92	90	13	94	100	46	81	94	100	94	100	94	96	94	81	94						
E. coli	84	42	99	36		93	93	93	93	82	88	94	88	96	83	100	89	100	88	93	50	77	80						
Klebsiella sp.	93	84	99	0		93	97	95	97	87	98	98	93	97	92	100	99	100	99	98	96	95	92						
Proteus sp.	40	82	100	52		92	100	100	100	88	70	100	92	100	85	100	85	100	57	100	82	100	88						
Pseudomonas sp.	289		85			78	41	93	28		81	86			75	91	83	94		95	53	81	89						
Serratia sp.	51	16	98	18		92	98	88	88	2	88	98	6	98	98	98	98	98	94	88	98	98	88						
Stenotrophomonas	83							29									87				98	39							

ORGANISM	# ISOLATES																												
		Amp/Sulbactam	Ampicillin	Augmentin	Azithromycin	Chloramphenicol	Ceftioxone	Clindamycin	Cephalexin	Cefaclor	Ceftaxime	Cefazolin	Ciprofloxacin	Cefepime	Cefturoxime	Daptomycin	Erythromycin	Ertapenem	Gentamicin	Gent/Syn	Imipenem	Levofloxacin	Linezolid	Meropenem	Moxifloxacin	Oxacillin	Penicillin	Pip/Tazo	Rifampin
<b>GRAM POSITIVE</b>		A/S	AM	AUG	AZI	C	CAX	CD	CF	CFR	CFT	CFZ	CP	CPE	CRM	DAP	E	ETP	GM	GMS	IMP	LVX	LZD	MER	MXF	OX	P	P/T	RIF
Enterococcus sp.	66		92									85				100	28			80		71	98				92		58
MRSA	383	0	0	0		86	0	73	0			0	49			99	8		98		0	52	99		76	0	0		97
MSSA	245	100	13	99		96	99	79	99			99	87			99	55	100	99		99	90	100		97	100	14	99	96
Coag Neg Staph	122	54	25	54			53	89				54	73			99	45	100	94			75	100			54	25		97
Strep pneumoniae	60			78	41	100	93	78		83	95			88	89		39					98		71			54		

\*17% of the MSSA were D test positive (Inducible Clindamycin Resistance)  
 14% of the Coag Neg Staph were D test positive (Inducible Clindamycin Resistance)  
 61% of the Staph aureus isolated were MRSA  
 Data collected 01/01/10 thru 12/31/10

**UT HEALTH NORTHEAST  
DEPARTMENT OF PATHOLOGY - MICROBIOLOGY  
2015 NON-URINE ANTIBIOGRAM**

ORGANISM	# ISOLATES	% SUSCEPTIBLE																				
		ANTIMICROBIC																				
GRAM NEGATIVE		A/S	AK	AM	AZT	CAX	CAZ	CFT	CFZ	CFZolin	CP	CPE	CRM	ETP	GM	IMP	LVX	MER	P/T	T/S	TGC	TO
Acinetobacter sp.	88	83	91			65	84	52			80	80			85		85	94			86	86
Achromobacter sp.	37		64		17	44	50	28			44	47			67	92	72	89	78	86		61
Enterobacter sp.	163	48	98	22	88	83	93	90	21	98	97	47	99	93	94	99	99	99	95	93	98	94
E. coli	135	45	99	40	93	93	93	94	83	64	94	91	100	84	99	64	100	100	97	59	100	84
Klebsiella sp.	210	81	99	<1	97	98	98	98	84	99	99	92	99	97	100	99	100	100	99	93	99	97
Proteus sp.	58	74	98	62	90	93	95	95	69	71	95	90	100	95	50	84	100	100	67	100	95	
Pseudomonas sp.	600		87		66	50	90	29		87	85			76	86	87	92	95	62		92	
Serratia sp.	162	15	99	8	69	60	65	70	<1	94	97	1	100	94	95	97	100	57	97	99	86	
Stenotrophomonas sp.	244						38									91				99		

ORGANISM	# ISOLATES	ANTIMICROBIC																													
		A/S	AM	AUG	AZI	C	CAX	CD	CFR	CFT	CFZ	CP	CPE	CRM	DAP	E	GM	GMS	IMP	LVX	LZD	MER	MXF	OX	P	RIF	STS	SYN	T/S	TE	VA
GRAM POSITIVE																															
Enterococcus sp.	60		95			0					70			97	12		77		75	98					95	45	70			17	95
MRSA	261	0		0		82	100	68		0	35			99	12	98		0	38	99	0	70	0	0	95		99	99	95	100	
MSSA	374	99	0	100		89	54	80		100	86			99	66	99		100	89	98	100	95	100	19	96		99	99	95	100	
Coag Neg Staph	155	54	0	54		98	68				65			97	38	88			66	99		78	54	17	99		98	64	82	99	
Strep pneumoniae	42			98	63	95		83	71	98			95	80	63				100		93			71					76	78	100

Data collected 01/01/15 through 12/31/15

41.0% of the Staph aureus isolated were MRSA

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MSSA = Methicillin Susceptible Staph aureus

# **Update: Antibiotic Stewardship,**

Joe Sartor, Pharm.D.

SO, YA  
MEAN WE MADE  
THIS MESS  
OURSELVES?

YEP, SON,  
WE HAVE MET  
THE ENEMY  
AND HE IS US.



CEPHALOSPORIN

VANCO  
MYCIN

TB

VRE

C. DIFFICILE

MRSA

CILLIN

MRSA

CLINDA  
MYCIN

AG-RES

VRE

AG-RES

1971  
WIL  
KELL

# Antimicrobial Stewardship

- Antimicrobial stewardship refers to coordinated interventions designed to improve and measure the appropriate use of antimicrobial agents by promoting the selection of the optimal antimicrobial drug regimen including dosing, duration of therapy, and route of administration.
- There will be national or coordinated legislative or regulatory mandates designed to optimize use of antimicrobial therapy through antimicrobial stewardship.
- Given the societal value of antimicrobials and their diminishing effectiveness due to antimicrobial resistance, IDSA supports broad implementation of antimicrobial stewardship programs across all health care settings



# Goals of Antimicrobial Stewardship

- To achieve best clinical outcomes by optimizing antimicrobial use
- Minimize toxicity and other adverse events
- Limit the selective pressure on bacterial populations that drives the emergence of antimicrobial-resistant strains
- Reduce health care associated infections
- Reduce the costs of inappropriate antimicrobial use

# The Joint Commission has announced a new Medication Management standard effective Jan. 1, 2017.

The elements of performance, in part, address:

- Leaders establishing antimicrobial stewardship as an organizational priority.
- Educating staff and licensed independent practitioners involved with ordering, dispensing, administering and monitoring antimicrobial resistance and stewardship practices.
- Educating patients and families on appropriate use of medications, including antibiotics.
- Creating a multidisciplinary, antimicrobial stewardship team.
- Developing an antimicrobial stewardship program.

# CMS

The antibiotic stewardship requirements in the final version of the CMS infection control survey include the following: COMMENT PERIOD ENDS 8/15/2016

- The hospital has written policies and procedures whose purpose is to improve antibiotic use (antibiotic stewardship).
- Designate leaders of the infection prevention and control program and the antibiotic stewardship program respectively, who are qualified through education, training, experience, or certification. This requirement allows for flexibility in staffing in order to suit the needs of each hospital or CAH.
- The hospital's antibiotic stewardship policy and procedures requires practitioners to document in the medical record or during order entry an indication for all antibiotics, in addition to other required elements such as dose and duration.
- The hospital has a formal procedure for all practitioners to review the appropriateness of any antibiotics prescribed after 48 hours from the initial orders (e.g., antibiotic time out).
- The hospital monitors antibiotic use (consumption) at the unit and/or hospital level.

# CDC Core Elements of Hospital Antibiotic Stewardship

- Leadership Commitment: Dedicating necessary human, financial and information technology resources
- Accountability: Appointing a single leader responsible for program outcomes. Experience with successful programs show that a physician leader is effective
- Drug Expertise: Appointing a single pharmacist leader responsible for working to improve antibiotic use.
- Action: Implementing at least one recommended action, such as systemic evaluation of ongoing treatment need after a set period of initial treatment (i.e. “antibiotic time out” after 48 hours)
- Tracking: Monitoring antibiotic prescribing and resistance patterns
- Reporting: Regular reporting information on antibiotic use and resistance to doctors, nurses and relevant staff
- Education: Educating clinicians about resistance and optimal prescribing

# CDC Key Support

- **Clinicians and department heads-** As the prescribers of antibiotics, it is vital that clinicians are fully engaged in and supportive of efforts to improve antibiotic use in hospitals.
- **Infection preventionists and hospital epidemiologists** coordinate facility-wide monitoring and prevention of healthcare-associated infections and can readily bring their skills to auditing, analyzing and reporting data.
- **Quality improvement staff** can also be key partners given that optimizing antibiotic use is a medical quality and patient safety issue.
- **Laboratory staff** can guide the proper use of tests and the flow of results. They can also guide empiric therapy by creating and interpreting a facility antibiogram. .
- **Information technology staff** are critical to integrating stewardship protocols into existing workflow.
- **Nurses** can assure that cultures are performed before starting antibiotics. In addition, nurses review medications as part of their routine duties and can prompt discussions of antibiotic treatment, indication, and duration.

# Interventions to improve antibiotic use

- Broad interventions
  - Antibiotic “Time outs”.
  - Prior authorization
  - Prospective audit and feedback
- Infection and syndrome specific interventions
  - Community-acquired pneumonia, Urinary tract infections (UTIs), Skin and soft tissue infections
  - Empiric coverage of methicillin-resistant *Staphylococcus aureus* (MRSA) infections
  - *Clostridium difficile* infections
  - Treatment of culture proven invasive infections

# Interventions to improve antibiotic use

- Pharmacy-driven Interventions
  - Automatic changes from intravenous to oral antibiotic therapy
  - Dose adjustments
  - Dose optimization
  - Automatic alerts in situations where therapy might be unnecessarily duplicative
  - Time-sensitive automatic stop orders
  - Detection and prevention of antibiotic-related drug-drug interactions

# Goals of Therapy Guidelines

- Use PK/PD of antimicrobials to promote the selection of the optimal antimicrobial drug regimen and minimize toxicity
- Decrease emergence of antimicrobial resistance
- Reviewed by Infectious Disease physician, Hospitalist, Intensivist, and Family Medicine physicians before presentation to PTCERC
- Promoted in Empiric Therapy order sets



# Antibiotic Stewardship Team

- Clinical Pharmacists
- ID Physicians
- Clinical Microbiologist
- Infection Control Specialist
- Meets weekly to review therapies for optimal utilization of antibiotics

# Antibiotic Stewardship

- Develop a formal, protocol-based, pharmacist-driven pharmacokinetic dosing program for antibiotics such as:
  - Vancomycin
  - Aminoglycosides
  - Time-dependent beta-lactam antibiotics
  - Antibiotic dosing requiring adjustments for renal/liver dysfunction

Pharmacokinetics and pharmacodynamics are interrelated such that, with respect to antimicrobials, they determine the relationship between serum drug concentrations and antimicrobial effect.

Pharmacokinetics is most important when determining dosing frequency, duration of infusion and effects on antimicrobial resistance

Different classes of antimicrobials have different pharmacodynamic properties.

# Pharmacokinetics and Pharmacodynamics

- Vancomycin has concentration dependent bactericidal activity
  - dosed renal to a targeted trough of 15mcg/ml (12mcg-17mcg/ml)
  - check trough every 4–7 doses or if significant SCr change
  - utilize MDRD6 to calculate eGFR to estimate trough
  - are looking at AUC/MIC ratios for  $\geq 85$ yo or poor renal function (eGFR < 20ml/minute)
- Aminoglycosides and fluoroquinolones have concentration-dependent bactericidal activity
  - higher the serum concentration, the greater the bactericidal activity of aminoglycosides Peak/MIC ratios
  - AUC/MIC ratio best estimate for fluoroquinolone activity

# Pharmacokinetics and Pharmacodynamics

- For beta-lactams the dose–response relationship is time dependent
  - The bactericidal activity is dependent on the time ( $t$ ) that the free drug concentration ( $f$ ) remains above the minimum inhibitory concentration (MIC) during the dosing interval ( $ft > MIC$ ).
- maximal efficacy occurs at a concentration four to five times higher than the MIC

Cr a i g WA . Pharmacokinetic /pharmacodynamic parameters: rationale for antibacterial dosing of mice and men.

*Clin Infect Dis.* 1998; 26:1-10.

Owens RC, Shorr AF. Rational dosing of antimicrobial agents: pharmacokinetic and pharmacodynamic strategies. *Am J Health-Syst Pharm.* 2009; 66(suppl 4):S23-30.

Arnold A, Brouse SD, Pitcher WD et al. Empiric therapy for gram-negative pathogens in nosocomial and health care associated pneumonia: starting with the end in mind. *J Intensive Care Med.* 2010;25:259-70.

Kim A, Sutherland CA, Kuti JL et al. Optimal dosing of piperacillin-tazobactam for the treatment of *Pseudomonas aeruginosa* infections: prolonged or continuous infusion? *Pharmacotherapy.* 2007;27:1490-7.

# Pharmacokinetics and Pharmacodynamics

Free beta-lactam concentrations do not have to remain above the MIC for the entire dosing interval. The percentage of time required for both bacteriostatic and maximal bactericidal activity is different for the various classes of beta-lactams. **Carbapenems** require free drug concentrations to exceed the MIC 20% of the dosing interval for bacteriostatic activity and 40% of the dosing interval for maximal bactericidal activity. **Cephalosporins** require free drug concentrations to be above the MIC for 35—40% of the dosing interval for bacteriostatic activity and 60—70% of the dosing interval for bactericidal activity. **Penicillins** require free drug concentrations to exceed the MIC for 30% of the dosing interval to achieve bacteriostatic activity and 50% of the dosing interval to achieve bactericidal activity.

DeRyke CA, Lee SY, Kuti JL, et al. Optimising dosing strategies of antibacterials utilising pharmacodynamic principles: impact on the development of resistance. *Drugs* 2006;66:1-14.

Drusano GL. Prevention of resistance: a goal for dose selection for antimicrobial agents. *Clin Infect Dis* 2003;36(S1):S42-50.

Lodise TP, Lomaestro BM, Drusano GL. Application of antimicrobial pharmacodynamic concepts into clinical practice: focus on beta-lactam antibiotics. *Pharmacotherapy* 2006;26:1320-1332.

# MIC Values

- **NOTE: MIC values vary from one drug to another and from one bacterium to another, and thus MIC values are NOT comparable between antibiotics or between organisms.**
- MIC values are used as indicators of appropriate therapies.

# Future

- Nursing homes
- Ambulatory practice
- Continuous infusion



**HAVE MORTAR WILL PESTLE**



**Joe Sartor, Pharm.D.**

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